

Epidemiology and Geographical Distribution of Human Schistosomiasis in Kenya: A Systematic Review

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ARTICLE INFORMATION	ABSTRACT
Article history: Published: September 2026 Keywords: Schistosomiasis Epidemiology Kenya Systematic Review Geographical Distribution	Schistosomiasis is a neglected tropical disease of significant public health importance in sub-Saharan Africa, ranking second in terms of years lived with disability. Despite numerous epidemiological studies conducted in Kenya, a comprehensive overview of the disease status nationally is lacking. This systematic review aimed to synthesize existing knowledge on the epidemiology and geographical distribution of human schistosomiasis in Kenya. Following PRISMA guidelines, we systematically searched PubMed, Web of Science, CAB Direct, and grey literature sources for records published between January 2000 and December 2020. Studies reporting prevalence data on human schistosomiasis conducted in Kenya were included. Study quality was assessed using Joanna Briggs Institute appraisal tools. Prevalence data were extracted and mapped using QGIS version 3.16.8. Of 1,343 initially retrieved records, 99 met inclusion criteria. <i>Schistosoma mansoni</i> and <i>S. haematobium</i> were the only species reported, with infections occurring across all age groups including preschool children, schoolaged children, and adults. <i>S. mansoni</i> was predominantly distributed in Western Kenya around Lake Victoria, while <i>S. haematobium</i> was concentrated along the Kenyan Coast. Spatiotemporal analysis revealed emergence of new transmission foci, with prevalence declining in some counties, increasing in others, and remaining stable in several areas. Hotspot counties included Homabay, Kirinyaga, Busia, Tana River, and Siaya. Key drivers of infection included geographical location, environmental factors, intermediate snail host distribution, human water contact patterns, and untreated at-risk populations. Schistosomiasis remains a public health challenge in Kenya. These findings highlight the need for expanded preventive chemotherapy to all at-risk populations, integrated with complementary interventions including health education, focal mollusciciding, and improved water, sanitation, and hygiene services.

1. Introduction

Schistosomiasis (bilharzia) is caused by trematode parasites of the genus *Schistosoma*, transmitted when humans come into contact with freshwater contaminated with cercariae released by infected intermediate snail hosts [1]. In Africa, *Schistosoma mansoni* and *Schistosoma haematobium* are the predominant species, causing intestinal and urogenital schistosomiasis respectively [2,3]. The disease is closely associated with poverty, poor sanitation, lack of clean water access, and limited health education [4,5].

Globally, schistosomiasis affects over 249 million people, with 97% of cases occurring in Africa [6]. The disease accounts for an estimated 2456 million disability-adjusted life-years (DALYs) lost globally [7], with approximately 3.3 million DALYs in sub-Saharan Africa [8]. In Kenya, approximately 6 million people are infected, with an estimated 17.4 million at risk of infection [9,10]. The Kenyan government, through the Ministries of Health and Education, launched a national school-based deworming program in 2009, incorporating praziquantel (PZQ) in 2012 for schistosomiasis control [11,12]. Mass drug administration (MDA) targeting schoolaged children (SAC) remains the primary control strategy, aligned with WHO's global schistosomiasis control roadmap [13,14].

Despite numerous epidemiological studies investigating schistosomiasis prevalence, distribution, and risk factors across Kenya [15,18], no comprehensive synthesis of this evidence exists. Following the adoption of WHO's Neglected Tropical Diseases roadmap 2021-2030 [19], which emphasizes disease elimination and expanded treatment coverage, understanding the current epidemiological landscape is crucial for guiding evidence-based control strategies. This systematic review aims to synthesize existing knowledge on the epidemiology and geographical distribution of human schistosomiasis in Kenya to inform effective, sustainable control strategies.

2. Methodology

2.1 Search Strategy

This systematic review followed the Preferred Reporting Items for Systematic Reviews and MetaAnalyses (PRISMA) guidelines [20]. Two independent reviewers systematically searched PubMed, Web of Science, CAB Direct, and grey literature sources (OpenGrey, OAIster) using the search phrase: (schistosomiasis OR *Schistosoma* OR Bilharzia OR "*S. mansoni*" OR "*S. haematobium*" OR "trematode infection") AND Kenya. Additionally, MSc/PhD theses from Kenyan university archives were searched using "schistosomiasis AND Kenya" in Google. A date restriction of January 2000 to December 2020 was applied.

2.2 Eligibility Criteria

Records were included if they: (i) investigated human schistosomiasis; (ii) were conducted in Kenya; (iii) were published between 2000-2020; (iv) were in English; (v) reported prevalence data with clearly stated geographical location. Exclusion criteria included duplicate records, studies with different scope (experimental studies, reviews, model predictions), studies on special populations (travelers, immigrants, refugees), and articles whose full text was unavailable even after author contact. Reference lists of included articles were manually searched for additional relevant records. Disagreements between reviewers were resolved through discussion and consensus with a third party.

2.3 Quality Assessment

Study quality was assessed using Joanna Briggs Institute (JBI) critical appraisal tools for prevalence studies [21] and cohort studies [22]. Each checklist item was scored 0 (not reported/unclear) or 1 (yes), with total scores ranging from 09 (prevalence studies) and 012 (cohort studies). Studies meeting $\geq 50\%$ of quality assessment criteria were included in the final analysis.

2.4 Data Extraction

From each included study, we extracted: author information, publication year, study period, study design, study setting, county/province, study population, age range, diagnostic method, *Schistosoma* species, number tested and positive, prevalence with 95% confidence intervals (CI), and morbidity markers where reported.

2.5 Data Analysis and Mapping

Data were categorized into two periods: preMDA (2000-2011) and postMDA (2012-2020). Prevalence and species distribution maps were generated using QGIS version 3.16.8. Prevalence was categorized according to WHO thresholds [13]: low ($<10\%$), moderate ($10-50\%$), and high ($>50\%$). "No information" was added for counties with no retrievable data. "Hotspots" were defined as areas with persistent high prevalence despite MDA. Confidence intervals were calculated in R version 3.4.2 using `binconf()` function from the `Hmisc` package. Graphical representations were created using Microsoft Excel.

3. Findings

3.1 Study Selection and Characteristics

The search yielded 1,343 records. After removing 262 duplicates, 880 records were excluded based on title and abstract screening. Fulltext assessment of 201 records led to exclusion of 102, resulting in 99 studies included in the final analysis (Figure 1). No additional records were identified through manual reference searching.

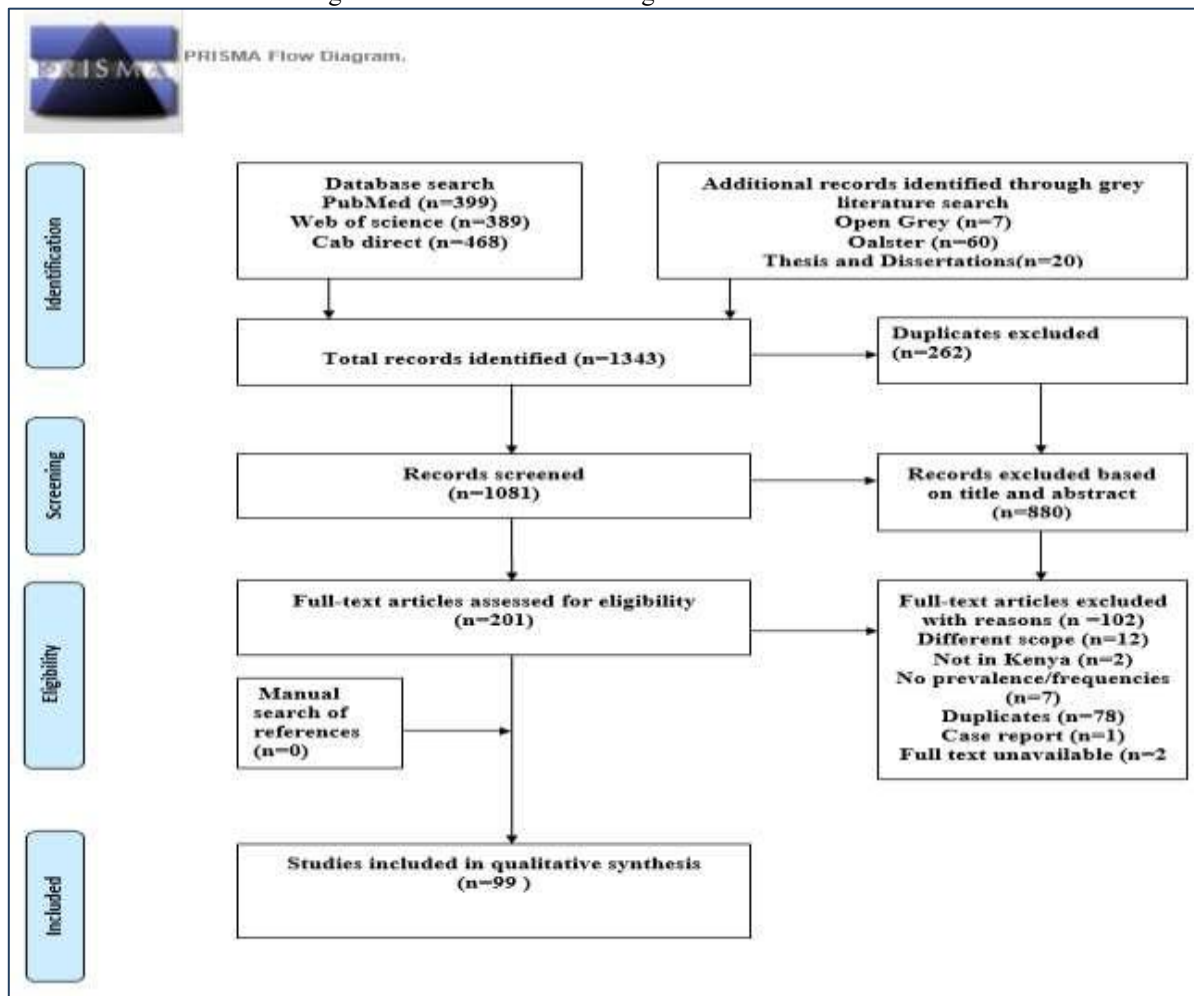


Figure 1. PRISMA flow diagram showing study selection process

Among included studies, 36 were published between 2000–2011 and 63 from 2012 onward. Study designs included cross-sectional (85.9%, $n=85$), longitudinal cohort (13.1%, $n=13$), and cluster-randomized trials (0.1%, $n=1$). Study settings comprised school-based (52, 55.5%), community-based (41, 44.4%), and hospital-based (3, 0.03%) studies. Records were distributed across all eight Kenyan provinces, with most from Nyanza (39.4%) and Coast (38.3%) provinces, while North Eastern (0.01%) and Rift Valley (0.01%) had the fewest reports. Participant age ranged from 197 years, with most studies (58.6%) conducted in SAC (518 years). *S. mansoni* was reported in 58 studies (58.6%), *S. haematobium* in 32 (32.3%), and mixed infections in 9 (9.1%). Sample sizes ranged from 36 to 29,619 participants. Microscopy (Kato-Katz, 67.1%; urine filtration, 41.8%) were the most common diagnostic methods (Figure 2).

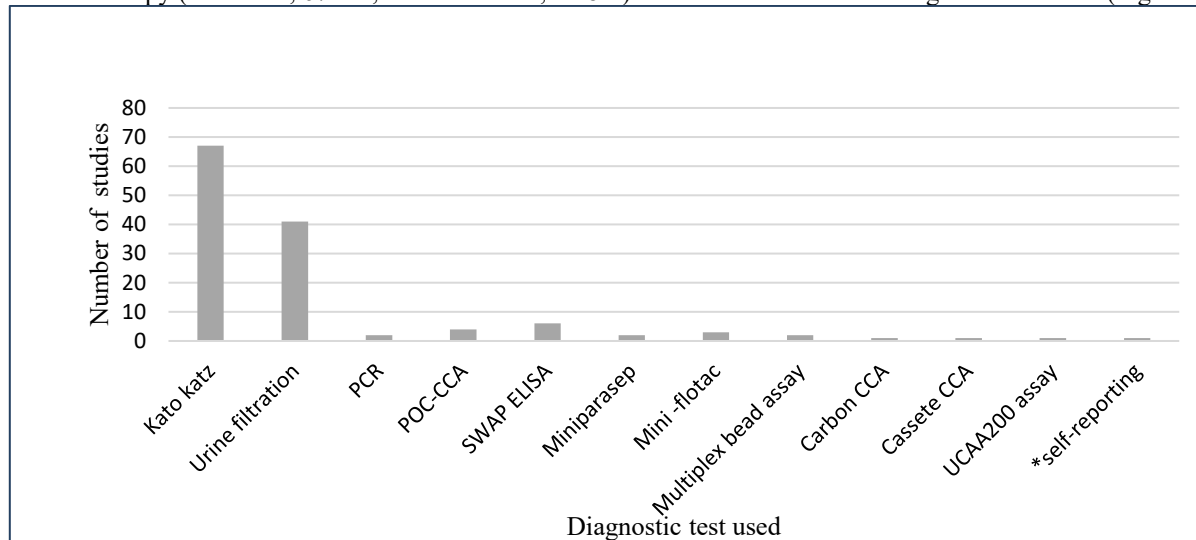


Figure 2. Diagnostic tests used in the studies included in this systematic review.

Figure 2. Diagnostic tests used in included studies. PCR: Polymerase Chain Reaction; POC-CCA: Point of Care Circulating Cathodic Antigen; ELISA: Enzyme Linked Immunosorbent Assay; CCA: Circulating Cathodic Antigen; *blood in urine

All age groups tested positive for *Schistosoma* infection, including preschool aged children (PSAC) [2326], SAC [10,2730], adults [3135], and pregnant women [36,37]. SAC showed the highest prevalence, reaching 94.3% for *S. haematobium* in Coast Province and 88.5% for *S. mansoni* in Nyanza Province [38,39]. Among PSAC, *S. mansoni* prevalence ranged from 2580.3% [23,24], while only one study reported *S. haematobium* prevalence of 2.9% in this age group [40]. In adults, *S. mansoni* prevalence ranged from 0.973.2% [41], with no *S. haematobium* infection reported.

Mixed *S. mansoni*/*S. haematobium* infections were documented exclusively in SAC [15,17,42,43]. Table 1 summarizes the characteristics of included studies.

Table 1. Summary of key characteristics of included schistosomiasis studies by province, Kenya (2000–2020)

Province	Studies (n, %)	Dominant Schistosome Species	Main Study Populations	Common Diagnostic Methods	Key Epidemiological Findings
Nyanza	39 (39.4%)	<i>S. mansoni</i> (highly endemic)	School-aged children (SAC), Pre-SAC, Adults	Kato-Katz, Urine filtration	Highest <i>S. mansoni</i> prevalence (up to 88.5%); intense transmission around Lake Victoria shores and islands; multiple hotspots (e.g., Homabay, Siaya).
Coast	38 (38.3%)	<i>S. haematobium</i> (highly endemic)	SAC, Adults, Pregnant women	Urine filtration, Kato-Katz	Highest <i>S. haematobium</i> prevalence (up to 94.3%); transmission linked to inland ponds, streams, and dams; persistent hotspots in Tana River and Kwale.
Western	5 (5.1%)	<i>S. mansoni</i>	SAC, Pre-SAC	Kato-Katz, POC-CCA	Endemic foci; notable transmission in Busia and Kakamega; moderate prevalence (e.g., 28.7% in Busia).
Eastern	5 (5.1%)	<i>S. mansoni</i>	SAC, Adults	Kato-Katz	Foci associated with irrigation schemes; historically high prevalence (e.g., Makueni ~88%) but significant declines noted in recent periods.
Central	5 (5.1%)	<i>S. mansoni</i>	SAC, Adults, Pre-SAC	Kato-Katz	Persistent hotspot in Kirinyaga (Mwea Irrigation Scheme); prevalence rebounded despite MDA (increased from <10% to >50% over time).
Rift Valley	2 (2.0%)	<i>S. mansoni</i>	SAC	Kato-Katz, POC-CCA	Low overall prevalence; scattered transmission foci (e.g., Bomet, Narok) with prevalence generally <5%.
Nairobi	2 (2.0%)	<i>S. mansoni</i>	SAC, Adults	Kato-Katz	Very low prevalence (<1%); minimal local transmission reported.

North Eastern	1 (1.0%)	<i>S. haematobium</i>	SAC	Urine filtration	Extremely low prevalence (0.5%); thermal exclusion likely limits snail hosts, though data are sparse.
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3.3 Geographical Distribution

Schistosomiasis was distributed across all eight provinces and 24 of 47 counties (Figure 3). In Nyanza Province, infection was concentrated along Lake Victoria shores, with higher prevalence on islands compared to mainland [44,45]. In Coast and Eastern provinces, transmission occurred around inland water bodies including ponds, streams, dams, and rivers. Scattered *S. mansoni* foci were identified in Central, Eastern, and Rift Valley provinces associated with irrigation schemes [10,46,47].

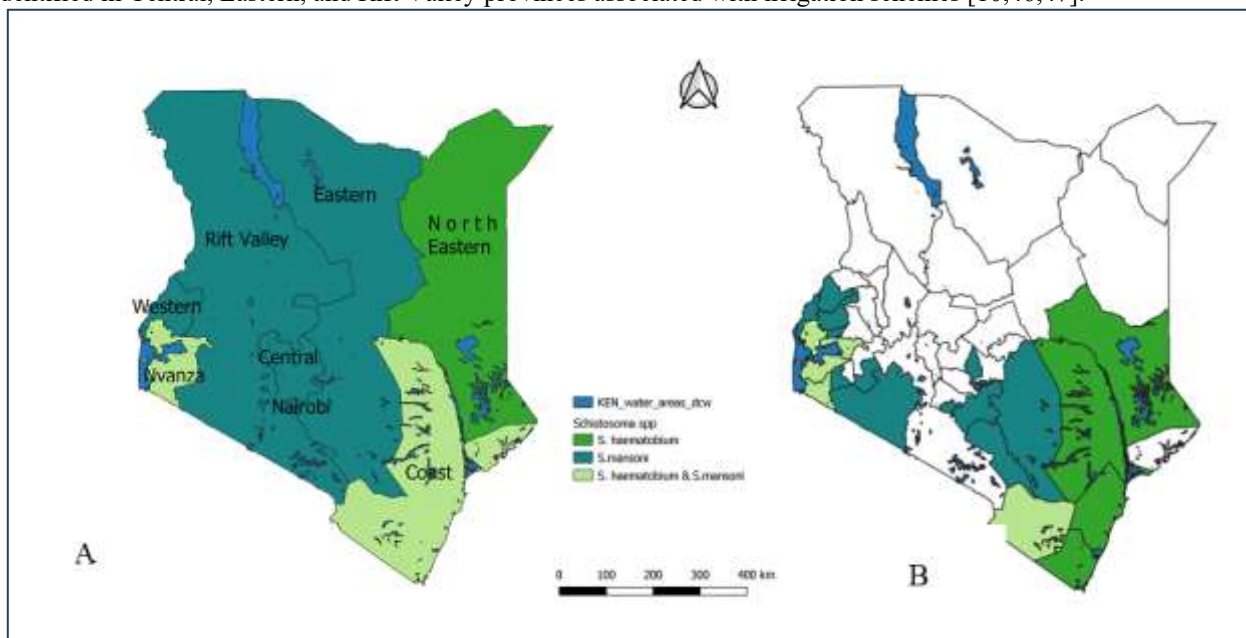


Figure 3. Geographical distribution of *Schistosoma* spp. infection in Kenya (A) by province and (B) by county

[Insert two maps: provincial distribution showing species patterns and county-level prevalence map]

S. haematobium predominated in North Eastern Province, while *S. mansoni* was reported in Western, Nairobi, Central, and Eastern provinces. Both species were documented in Nyanza and Coast provinces. A positive correlation between infection prevalence and proximity to water sources was noted across multiple studies [18,42,48,49].

3.4 SpatioTemporal Patterns

Prevalence among SAC showed significant changes between 2000/2011 and 2012/2020 (Figure 4). In Machakos and Makueni counties (Eastern Province) and Kilifi County (Coast Province), prevalence declined from >50% to <10%. Conversely, Kirinyaga County (Central Province) transitioned from low risk (<10%) to high risk (>50%) in the second period [50]. Homabay County (Nyanza Province) emerged as a new high risk area with prevalence >50% [51,52], despite being nonendemic in the earlier period.

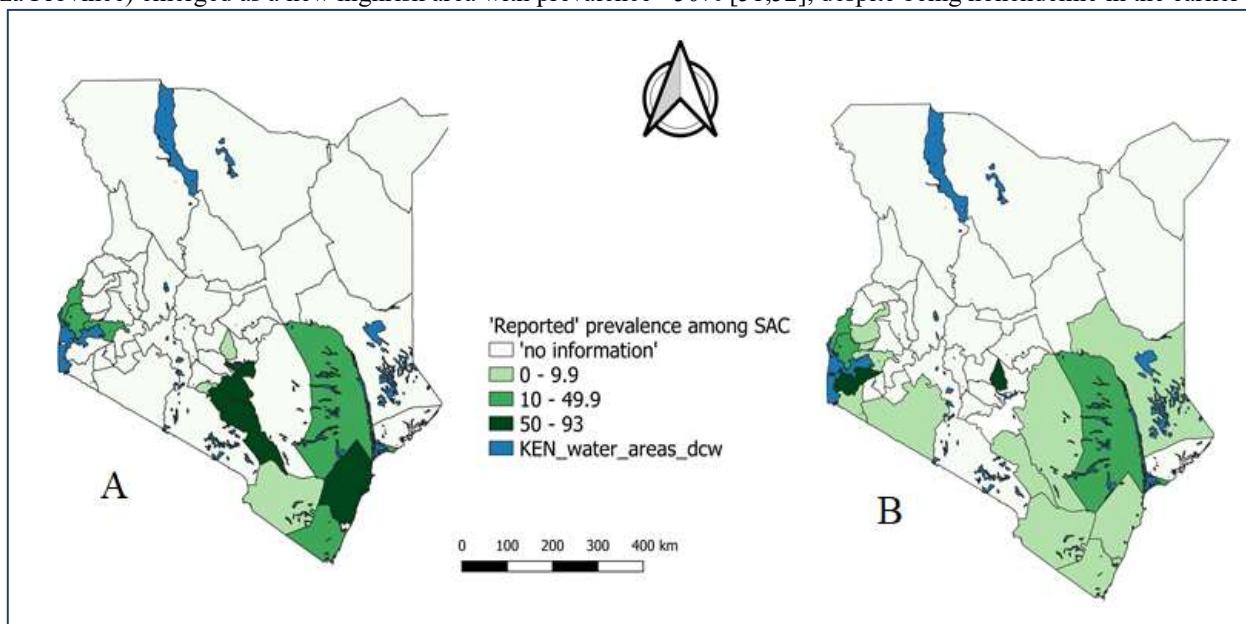


Figure 4. Spatiotemporal changes in *Schistosoma* spp. prevalence among SAC (A) 2000/2011 and (B) 2012/2020

Persistent hotspots included Tana River, Siaya, and Busia counties, with stable high prevalence across both periods. Specific transmission hotspots identified included Budalangi School (Busia County) [53], Mianya School (Kirinyaga County) [10,54,55], Dunga and Kaenyika Schools (Homabay County) [56], and Luanda K'Otieno beach (Siaya County) [57].

3.5 Schistosomiasis Associated Morbidity

Only 18.2% (18/99) of studies reported schistosomiasis associated morbidity, primarily conducted in SAC [36,58,59]. Reported morbidities included hepatomegaly (666.9%), splenomegaly (29.256.3%), hepatosplenomegaly (951.1%), anemia (19.354.7%), macrohematuria (9.568.1%), urinary tract abnormalities (1784%), wasting (710%), and stunting (20.532.5%) (Table 2). Assessment methods included ultrasonography for organopathology, hemoglobin measurement for anemia, urine dipstick for hematuria, and anthropometric measurements for nutritional status.

Table 2. Schistosomiasis's associated morbidity markers retrieved from included studies.

Study Period	Study design	Study group	Hepatomegaly (%)	Splenomegaly P (%)	HS (%)	Anemia P (%)	UT abnormalities (%)	Macro-hematuria (%)	Wasting and, stunting (%)	Ref
2000-2011	CS	SAC	—	—	51.1	—	17	—	—	[130]
2000-2011	CS	All-age	—	—	—	—	54	—	—	[68]
2000-2011	CS	All-age	—	—	—	—	14	51	—	[100]
2000-2011	CS	All-age	—	—	—	—	4	—	—	[90]
200-2011	Cohort	adults	—	—	—	—	5	39	—	[67]
2000-2011	Cohort	SAC	6	50.7	—	—	—	—	—	[71]
2000-2011	Cohort	SAC	6	36	9	—	—	—	—	[78]
2012-2020	Cohort	SAC	66.9	56.3	—	19.3-20.5	47.1	—	7-10.8 & 20.5-32.5	[43]
2012-2020	CS	SAC	—	—	—	—	84	—	—	[40]
2012-2020	CS	SAC	—	—	—	—	41	—	—	[60]
2012-2020	CS	SAC	—	—	—	54.7	—	—	—	[47]
2012-2020	CS	PSAC	50	29.2	—	—	—	—	—	[64]
2012-2020	Cohort	Adults	—	—	—	71	—	—	—	[85]
2012-2020	CS	Adults	—	—	—	—	17	—	—	[54]
2012-2020	CS	SAC	—	—	—	33.9	—	33.8	—	[94]
2012-2020	CS	All-age	—	—	—	—	—	13.5	—	[105]
2012-2020	CS	SAC	—	—	—	25.7	—	—	—	[35]
2012-2020	CS	All age	—	—	—	—	—	68.1	—	[82]

P: prevalence, UT: Urinary tract, CS: Cross-sectional studies, HS; Hepatosplenomegaly; Ref: Reference.

4. Discussion

This is the first systematic review synthesizing the epidemiology and distribution of human schistosomiasis in Kenya over a 20year period (2000-2020). Our findings demonstrate that both *S. mansoni* and *S. haematobium* remain prevalent across Kenya, with transmission occurring in all age groups, reflecting ongoing exposure and active transmission cycles.

4.1 Age Specific Infection Patterns

The high prevalence observed in SAC aligns with findings from other African countries [60,61] and reflects the increased water contact patterns and behavioral factors in this age group. The documented infection in PSAC [23,25,26,40] underscores the need for expanding treatment coverage to younger children, as recommended in a recent subSaharan Africa systematic review [62]. Adults, including pregnant women, also maintain significant infection prevalence [34,36,63], acting as reservoirs sustaining community transmission. These findings challenge the current school-focused MDA approach and support calls for communitywide treatment strategies [64].

4.2 Diagnostic Method Trends

While KatoKatz and urine filtration remained the predominant diagnostic methods throughout the study period, there was increased utilization of more sensitive tests including PointofCare Circulating Cathodic Antigen (POCCCA) and PCR in the second decade (2012-2020). This shift likely reflects WHO's recommendation for improved diagnostics as prevalence declines with intensified interventions [13]. The use of more sensitive methods may partly explain the emergence of new foci and detection of low intensity infections previously missed by microscopy [65,66].

4.3 Morbidity Assessment

The increased reporting of morbidity markers in the second study period corresponds with WHO recommendations to assess chemotherapy impact using morbidity indicators [67,68]. SAC bear the highest morbidity burden, correlating with their higher infection prevalence and intensity. The lower morbidity observed in adults likely reflects acquired immunity and benefits of early antischistosomal treatment [69]. These findings emphasize the need for continued morbidity monitoring to evaluate control program effectiveness.

4.4 Geographical Distribution Drivers

The heterogeneous distribution of schistosomiasis in Kenya reflects the ecology of intermediate snail hosts and human water contact patterns. The Lake Victoria basin's high *S. mansoni* prevalence is associated with abundant *Biomphalaria* snails, favorable climatic conditions, and communities' reliance on lake water [18,44,45,70]. Similarly, *S. haematobium* predominance along the Coast

correlates with Bulinus snail distribution in inland water bodies with suitable temperatures and currents [71,72]. The overlap of both species in Coast and Lake Victoria regions indicates risk of coinfections in predisposed populations [15,17,42,43]. The limited data from North Eastern Province may reflect either minimal transmission due to thermal exclusion of snail hosts [71,73,74] or underinvestigation of these areas. This pattern mirrors observations from northern Uganda [75].

4.5 SpatioTemporal Dynamics and Hotspots

The changing prevalence patterns over 20 years reveal important epidemiological transitions. Declining prevalence in some counties likely reflects positive impacts of MDA combined with other control measures. However, the emergence of new foci (Homabay) and increasing prevalence in previously lowrisk areas (Kirinyaga) suggest ongoing transmission dynamics. Several factors may explain these changes: population migration from endemic areas, use of more sensitive diagnostics, lack of improved water sources [46], poor hygiene practices [43,48,49], high population density [43], waterway obstruction increasing snail habitats [76], and untreated populations perpetuating transmission [46,50,63].

Persistent hotspots demonstrate ongoing high transmission despite interventions. In Mwea County, where MDA was first piloted but implemented without complementary health education and WASH interventions, prevalence rebounded over time [54]. This underscores the limitation of chemotherapy alone and the critical need for integrated approaches [10,55,77].

4.6 Implications for Control

WHO recommends preventive chemotherapy for all atrisk populations in hotspot areas, combined with complementary interventions [13]. Our findings support this recommendation, particularly regarding expansion beyond SAC. Evidence from Kenya demonstrates that communitybased approaches can be effective for mass treatment [78], while combining chemotherapy with intermittent mollusciciding provides longerterm benefits than chemotherapy alone [79].

4.7 Study Limitations

Several limitations should be considered. First, variation in diagnostic methods may have led to under or overestimation of prevalence. Second, despite studies from all provinces, data were not available for all counties, with some provinces underrepresented. Third, data heterogeneity in terms of province, study groups, design, and setting precluded metaanalysis for pooled prevalence estimates. Nonetheless, this review's strengths include the comprehensive search across three major databases and grey literature, inclusion of 99 goodquality studies, and adherence to PRISMA guidelines, ensuring robust synthesis.

5. Conclusion and Recommendations

Schistosomiasis remains a significant public health problem in Kenya, with both *S. mansoni* and *S. haematobium* infections occurring across all age groups. The disease is geographically heterogeneous, with *S. mansoni* predominating in the Lake Victoria region and *S. haematobium* along the Coast. Over the past 20 years, spatiotemporal patterns have shown emerging foci, persistent hotspots, and variable trends in prevalence. Key transmission drivers include geographical location, environmental factors, intermediate snail distribution, human water contact patterns, and untreated atrisk populations. These findings provide critical evidence for policy discussions and the design of effective, sustainable control strategies aligned with the 20212030 NTD roadmap. The authors recommend: (1) expanding preventive chemotherapy to all age groups in highrisk areas; (2) integrating schoolbased programs with communitydirected interventions; (3) implementing focal mollusciciding in hotspot areas; (4) promoting health education to reduce water contact behaviors; and (5) improving water, sanitation, and hygiene (WASH) infrastructure.

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References

- [1] World Health Organization. Progress report 20012011 and strategic plan 20122020. Geneva: WHO; 2010.
- [2] Gryseels B, Polman K, Clerinx J, Kestens L. Human schistosomiasis. *Lancet*. 2006;368:110618.
- [3] Colley DG, Bustinduy AL, Secor WE, King CH. Human schistosomiasis. *Lancet*. 2014;383:225364.
- [4] Mnkugwe RH, Minzi OS, Kinung'hi SM, et al. Prevalence and correlates of intestinal schistosomiasis infection among schoolaged children in NorthWestern Tanzania. *PLoS One*. 2020;15:e0228770.
- [5] Ross A, Inobaya M, Olveda R, et al. Prevention and control of schistosomiasis: a current perspective. *Res Rep Trop Med*. 2014;5:65.
- [6] Steinmann P, Keiser J, Bos R, et al. Schistosomiasis and water resources development: systematic review, metaanalysis, and estimates of people at risk. *Lancet Infect Dis*. 2006;6:41125.
- [7] King CH, DangerfieldCha M. The unacknowledged impact of chronic schistosomiasis. *Chronic Illn*. 2008;4:6579.
- [8] Murray CJL, Vos T, Lozano R, et al. Disabilityadjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 19902010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*. 2012;380:2197223.
- [9] Masaku J, Madigu N, Okoyo C, Njenga SM. Current status of *Schistosoma mansoni* and the factors associated with infection two years following mass drug administration programme among primary school children in Mwea irrigation scheme: a crosssectional study. *BMC Public Health*. 2015;15:19.
- [10] Okoyo C, Campbell SJ, Williams K, et al. Prevalence, intensity and associated risk factors of soiltransmitted helminth and schistosome infections in Kenya: Impact assessment after five rounds of mass drug administration in Kenya. *PLoS Negl Trop Dis*. 2020;14:e0008604.
- [11] Ministry of Health, Kenya. Kenya National SchoolBased Deworming Programme. Nairobi: Government of Kenya; 2014.
- [12] Okoyo C, Nikolay B, Kihara J, et al. Monitoring the impact of a national school based deworming programme on soiltransmitted helminths in Kenya: the first three years, 20122014. *Parasit Vectors*. 2016;9:113.

- [13] World Health Organization. Ending the neglect to attain the Sustainable Development Goals: A road map for neglected tropical diseases 20212030. Geneva: WHO; 2020.
- [14] Savioli L, Stansfield S, Bundy DAP, et al. Schistosomiasis and soiltransmitted helminth infections: forging control efforts. *Trans R Soc Trop Med Hyg.* 2002;96:5779.
- [15] Ng'ang'a M, Matendechero S, Kariuki L, et al. Spatial distribution and coinfection with urogenital and intestinal schistosomiasis among primary school children in Migori County, Kenya. *East Afr Med J.* 2016;93:S2231.
- [16] Masaku J, Madigu N, Okoyo C, Njenga SM. Current status of *Schistosoma mansoni* and the factors associated with infection two years following mass drug administration programme among primary school children in Mwea irrigation scheme: a crosssectional study. *BMC Public Health.* 2015;15.
- [17] Amollo DA, Kihara JH, Kombe Y, Karanja SM. Prevalence and intensity of single and mixed *Schistosoma mansoni* and *Schistosoma haematobium* infections in primary school children in Rachuonyo North District, Homabay County, western Kenya. *East Afr Med J.* 2013;90:3644.
- [18] Sang HC, Muchiri G, Ombok M, et al. *Schistosoma haematobium* hotspots in south Nyanza, western Kenya: prevalence, distribution and coendemicity with *Schistosoma mansoni* and soiltransmitted helminths. *Parasit Vectors.* 2014;7:112.
- [19] World Health Organization. Ending the neglect to attain the Sustainable Development Goals. Geneva: WHO; 2020.
- [20] Liberati A, Altman DG, Tetzlaff J, et al. The PRISMA statement for reporting systematic reviews and metaanalyses of studies that evaluate health care interventions: explanation and elaboration. *PLoS Med.* 2009;6:e1000100.
- [21] Joanna Briggs Institute. Checklist for Prevalence Studies. Adelaide: JBI; 2017.
- [22] Joanna Briggs Institute. Checklist for Cohort Studies. Adelaide: JBI; 2017.
- [23] Arnold BF, Kanyi H, Njenga SM, et al. Finescale heterogeneity in *Schistosoma mansoni* force of infection measured through antibody response. *Proc Natl Acad Sci USA.* 2020;117:2317481.
- [24] Davis SM, Wiegand RE, Mulama F, et al. Morbidity associated with schistosomiasis before and after treatment in young children in Rusinga Island, western Kenya. *Am J Trop Med Hyg.* 2015;92:9528.
- [25] Kimani BW, Mbugua AK, Kihara JH, et al. Safety, efficacy and acceptability of praziquantel in the treatment of *Schistosoma haematobium* in preschool children of Kwale County, Kenya. *PLoS One.* 2018;9:112.
- [26] Verani JR, Abudho B, Montgomery SP, et al. Schistosomiasis among young children in Usoma, Kenya. *Am J Trop Med Hyg.* 2011;84:78791.
- [27] Chadeka EA, Nagi S, Sunahara T, et al. Spatial distribution and risk factors of *Schistosoma haematobium* and hookworm infections among schoolchildren in Kwale, Kenya. *PLoS Negl Trop Dis.* 2017;11:e0005872.
- [28] Kiiti RW, Omukunda EN, Korir JC. Risk factors associated with helminthic intestinal infection in Lurambi Subcounty, Kakamega, Kenya. *J Parasitol Res.* 2020;2020:8810519.
- [29] Mwandawiro C, Okoyo C, Kihara J, et al. Results of a national schoolbased deworming programme on soiltransmitted helminths infections and schistosomiasis in Kenya: 20122017. *Parasit Vectors.* 2019;12.
- [30] Njaanake KH, Vennervald BJ, Simonsen PE, et al. *Schistosoma haematobium* and soiltransmitted helminths in Tana Delta District of Kenya: infection and morbidity patterns in primary schoolchildren from two isolated villages. *BMC Infect Dis.* 2016;16:110.
- [31] Gichuki PM, Kepha S, Mulewa D, et al. Association between *Schistosoma mansoni* infection and access to improved water and sanitation facilities in Mwea, Kirinyaga County, Kenya. *BMC Infect Dis.* 2019;19:114.
- [32] Masaku J, Mutungi F, Gichuki PM, et al. High prevalence of helminths infection and associated risk factors among adults living in a rural setting, central Kenya: a crosssectional study. *Trop Med Health.* 2017;45:19.
- [33] Riner DK, Ndombi EM, Carter JM, et al. *Schistosoma mansoni* infection can jeopardize the duration of protective levels of antibody responses to immunizations against hepatitis B and tetanus toxoid. *PLoS Negl Trop Dis.* 2016;10:e0005180.
- [34] Won KY, Abudho B, Blackstock AJ, et al. Assessment of quality of life as a tool for measuring morbidity due to *Schistosoma mansoni* infection and the impact of treatment. *Am J Trop Med Hyg.* 2014;90:3228.
- [35] Andereck JW, Kipp AM, Ondiek M, Vermund SH. Helminth prevalence among adults in rural Kenya: a stool survey for soiltransmitted helminths and schistosomiasis in Nyanza province. *Trans R Soc Trop Med Hyg.* 2014;108:8049.
- [36] Kihara JH, Kutima HL, Ouma J, et al. Urogenital schistosomiasis in women of reproductive age and pregnant mothers in Kwale County, Kenya. *J Helminthol.* 2015;89:10511.
- [37] Kariuki H, Mbugua G, Magak P, et al. Prevalence and familial aggregation of schistosomal liver morbidity in Kenya: evaluation by new ultrasound criteria. *J Infect Dis.* 2001;183:9606.
- [38] Vennervald BJ, Kenty LC, Butterworth AE, et al. Detailed clinical and ultrasound examination of children and adolescents in a *Schistosoma mansoni* endemic area in Kenya: hepatosplenic disease in the absence of portal fibrosis. *Trop Med Int Health.* 2004;9:46170.
- [39] Njaanake KH, Simonsen PE, Vennervald BJ, et al. Urinary cytokines in *Schistosoma haematobium*infected schoolchildren from Tana Delta District of Kenya. *BMC Infect Dis.* 2014;14:18.
- [40] LaBeaud AD, Nayakwadi Singer M, McKibben M, et al. Parasitism in children aged three years and under: relationship between infection and growth in rural coastal Kenya. *PLoS Negl Trop Dis.* 2015;9:e0003721.
- [41] Andereck JW, Kipp AM, Ondiek M, Vermund SH. Helminth prevalence among adults in rural Kenya: a stool survey for soiltransmitted helminths and schistosomiasis in Nyanza province. *Trans R Soc Trop Med Hyg.* 2014;108:8049.
- [42] Nagi S, Chadeka EA, Sunahara T, et al. Risk factors and spatial distribution of *Schistosoma mansoni* infection among primary school children in Mbita District, Western Kenya. *PLoS Negl Trop Dis.* 2014;8:e0002991.
- [43] Chadeka EA, Nagi S, Cheruiyot NB, et al. A highintensity cluster of *Schistosoma mansoni* infection around Mbita causeway, western Kenya: a confirmatory crosssectional survey. *Trop Med Health.* 2019;47:14.
- [44] Odiere MR, Rawago FO, Ombok M, et al. High prevalence of schistosomiasis in Mbita and its adjacent islands of Lake Victoria, western Kenya. *Parasit Vectors.* 2012;5:29.

- [45] Ng'ang'a M, Matendecheo S, Kariuki L, et al. Spatial distribution and coinfection with urogenital and intestinal schistosomiasis among primary school children in Migori County, Kenya. *East Afr Med J*. 2016;93:S2231.
- [46] Gichuki PM, Kepha S, Mulewa D, et al. Association between *Schistosoma mansoni* infection and access to improved water and sanitation facilities in Mwea, Kirinyaga County, Kenya. *BMC Infect Dis*. 2019;19:114.
- [47] Kihara JH, Muhoho N, Njomo D, et al. Drug efficacy of praziquantel and albendazole in school children in Mwea Division, Central Province, Kenya. *Acta Trop*. 2007;102:16571.
- [48] Woodhall DM, Wiegand RE, Wellman M, et al. Use of geospatial modeling to predict *Schistosoma mansoni* prevalence in Nyanza Province, Kenya. *PLoS One*. 2013;8:e71635.
- [49] Takeuchi R, Njenga SM, Ichinose Y, et al. Is there a gap between health education content and practice toward schistosomiasis prevention among schoolchildren along the shores of Lake Victoria in Kenya? *PLoS Negl Trop Dis*. 2019;13:116.
- [50] Njambi E, Magu D, Masaku J, et al. Prevalence of intestinal parasitic infections and associated water, sanitation, and hygiene risk factors among school children in Mwea Irrigation Scheme, Kirinyaga County, Kenya. *J Trop Med*. 2020;2020:3974156.
- [51] Chadeka EA, Nagi S, Sunahara T, et al. Spatial distribution and risk factors of *Schistosoma haematobium* and hookworm infections among schoolchildren in Kwale, Kenya. *PLoS Negl Trop Dis*. 2017;11:e0005872.
- [52] Davis SM, Wiegand RE, Mulama F, et al. Morbidity associated with schistosomiasis before and after treatment in young children in Rusinga Island, Western Kenya. *Am J Trop Med Hyg*. 2015;92:9528.
- [53] Wanjala PM, Khaemba BM, Luvai AI. Prevalence and intensity of infection of intestinal schistosomiasis and reinfection after intervention in Budalangi endemic focus of Western Kenya. *Int J Trop Med*. 2013;8:7180.
- [54] Masaku J, Madigu N, Okoyo C, Njenga SM. Current status of *Schistosoma mansoni* and the factors associated with infection two years following mass drug administration programme among primary school children in Mwea irrigation scheme: a crosssectional study. *BMC Public Health*. 2015;15.
- [55] Kingori WI. The epidemiology and control of schistosomiasis and soil transmitted helminthes in two communities within the Mwea Irrigation Scheme in Central Kenya [MSc thesis]. Nairobi: Kenyatta University; 2008.
- [56] Odiero MR, Opisa S, Odhiambo G, et al. Geographical distribution of schistosomiasis and soiltransmitted helminths among school children in informal settlements in Kisumu City, Western Kenya. *Parasitology*. 2011;138:156977.
- [57] Zhang K. Bridging the gap on schistosomiasis: a crosssectional study examining the knowledge gap and common attitudes and practices regarding *S. mansoni* infections among varying education levels in Luanda K'Otieno, western Kenya [MSc thesis]. Nairobi: University of Nairobi; 2019.
- [58] Sircar AD, Mwinzi PNM, Onkanga IO, et al. *Schistosoma mansoni* mass drug administration regimens and their effect on morbidity among schoolchildren over a 5year periodKenya, 20102015. *Am J Trop Med Hyg*. 2018;99:3629.
- [59] Won KY, Kanyi HM, Mwendu FM, et al. Multiplex serologic assessment of schistosomiasis in Western Kenya: antibody responses in preschool aged children as a measure of reduced transmission. *Am J Trop Med Hyg*. 2017;96:14607.
- [60] Aula OP, McManus DP, Jones MK, Gordon CA. Schistosomiasis with a focus on Africa. *Trop Med Infect Dis*. 2021;6:109.
- [61] Mazigo HD, Uisso C, Kazyoba P, et al. Prevalence, infection intensity and geographical distribution of schistosomiasis among preschool and school aged children in villages surrounding Lake Nyasa, Tanzania. *Sci Rep*. 2021;11:111.
- [62] Kalinda C, Mindu T, Chimbari MJ. A systematic review and metaanalysis quantifying schistosomiasis infection burden in preschool aged children (PreSAC) in subSaharan Africa for the period 20002020. *PLoS One*. 2020;15:e0244695.
- [63] Njenga SM, Mwandawiro CS, Muniu E, et al. Adult population as potential reservoir of NTD infections in rural villages of Kwale district, Coastal Kenya: implications for preventive chemotherapy interventions policy. *Parasit Vectors*. 2011;4:175.
- [64] Mwinzi PNM, Montgomery SP, Owaga CO, et al. Integrated communitydirected intervention for schistosomiasis and soil transmitted helminths in western Kenya a pilot study. *Parasit Vectors*. 2012;5:110.
- [65] Meurs L, Brienens E, Mbow M, et al. Is PCR the next reference standard for the diagnosis of schistosoma in stool? A comparison with microscopy in Senegal and Kenya. *PLoS Negl Trop Dis*. 2015;9:e0003959.
- [66] Okoyo C, Simiyu E, Njenga SM, Mwandawiro C. Comparing the performance of circulating cathodic antigen and KatoKatz techniques in evaluating *Schistosoma mansoni* infection in areas with low prevalence in selected counties of Kenya: a crosssectional study. *BMC Infect Dis*. 2018;18:17.
- [67] World Health Organization. Prevention and control of schistosomiasis and soiltransmitted helminthiasis. Geneva: WHO; 2002.
- [68] Andrade G, Bertsch DJ, Gazzinelli A, King CH. Decline in infectionrelated morbidities following drugmediated reductions in the intensity of *Schistosoma* infection: a systematic review and metaanalysis. *PLoS Negl Trop Dis*. 2017;11:e0005372.
- [69] Ouma JH, King CH, Muchiri EM, et al. Late benefits 1018 years after drug therapy for infection with *Schistosoma haematobium* in Kwale District, Coast Province, Kenya. *Am J Trop Med Hyg*. 2005;73:35964.
- [70] Sang HC, Muchiri G, Ombok M, et al. *Schistosoma haematobium* hotspots in south Nyanza, western Kenya: prevalence, distribution and coendemicity with *Schistosoma mansoni* and soiltransmitted helminths. *Parasit Vectors*. 2014;7:112.
- [71] Kariuki HC, Clennon JA, Brady MS, et al. Distribution patterns and cercarial shedding of *Bulinus nasutus* and other snails in the Msambweni area, Coast Province, Kenya. *Am J Trop Med Hyg*. 2004;70:44956.
- [72] Clennon JA, Mungai PL, Muchiri EM, et al. Spatial and temporal variations in local transmission of *Schistosoma haematobium* in Msambweni, Kenya. *Am J Trop Med Hyg*. 2006;75:103441.
- [73] Brooker S, Kabatereine NB, Smith JL, et al. An updated atlas of human helminth infections: the example of East Africa. *Int J Health Geogr*. 2009;8:111.
- [74] Clennon JA, King CH, Muchiri EM, et al. Spatial patterns of urinary schistosomiasis infection in a highly endemic area of coastal Kenya. *Am J Trop Med Hyg*. 2004;70:4438.

- [75] Stensgaard AS, Jørgensen A, Kabatereine NB, et al. Modeling freshwater snail habitat suitability and areas of potential snailborne disease transmission in Uganda. *Geospat Health*. 2006;1:93104.
- [76] Wanjiku MR. Occurrence of *Schistosoma mansoni* and its transmission risks in Mwea Irrigation Scheme, Kirinyaga County, Kenya [MSc thesis]. Nairobi: Kenyatta University; 2015.
- [77] Mutahi WT. Prevalence and intensity of schistosomiasis mansoni in irrigation and nonirrigation areas of Central Kenya [MSc thesis]. Nairobi: Kenyatta University; 2005.
- [78] Mwinzi PNM, Montgomery SP, Owaga CO, et al. Integrated communitydirected intervention for schistosomiasis and soil transmitted helminths in western Kenya a pilot study. *Parasit Vectors*. 2012;5:110.
- [79] Kariuki HC, Madsen H, Ouma JH, et al. Long term study on the effect of mollusciciding with niclosamide in stream habitats on the transmission of schistosomiasis mansoni after communitybased chemotherapy in Makuani District, Kenya. *Parasit Vectors*. 2013;6:111.